Protocol: Basic Media (Yeast)

Basic medium is the base for synthetic complete and drop-out yeast medium. It is also the medium used for dissolving yeast media additives, like amino acids and glucose solutions.

The following reagents are going to be required to make liquid and agar synthetic media:

Reagents
Yeast Nitrogen Base -AA / -AS (note that this means lacking amino acids and ammonium sulfate)
Ammonium Sulfate $(NH_4)_2SO_4$
diH_2O

<u>Note:</u> When autoclaving liquid, the rule of thumb is to make sure the final volume of your liquid is no more than ½ the max volume of the container (e.g., no more than 500 ml of liquid in a 1 L Pyrex bottle.)

1. To an autoclavable bottle, add:

Reagent	**Amount to add per 500ml final volume**
Yeast Nitrogen Base (-AA / -AS)	0.85 g
Ammonium Sulfate $(NH_4)_2SO_4$	2.5 g
diH_2O	500 ml

2. Autoclave the mixture **(You <u>MUST</u> get trained on how to use the autoclave before doing this!)**
    a. Make sure the screw-caps are **<u>LOOSLEY</u>** screwed on, allowing gas to escape the bottle
    b. Place a piece of autoclave tape across the cap, making sure one end is taped to the glass bottle
    c. Autoclave for 25 minutes (use “SLOW EXHAUST” setting)

3. Allow the media to cool to room temperature before tightening the cap on the media for storage.

Note: I find that nothing grows in Basic Medium. Once bottles are cool, I have combined two 500 ml volumes into a single 1L flask simply by pouring one into the other. Combining volumes allows us to minimize space in the media cabinet and wash one of the two bottles for future use.